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GB 5009.259-2016

National food safety standard Determination of biotin in foods

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National food safety standard

Determination of Biotin in foods

1 Scope

This standard specifies the method for determination of biotin in food.

This standard applies to the determination of biotin in food.

2 Principles

Biotin is a nutrient necessary for the growth of *Lactobacillus plantarum*. In the biotin determination medium, the growth of *Lactobacillus plantarum* is in linear relation with the biotin content in the sample to be determined; AND the content of the substance to be determined in the sample is calculated by comparing the light transmittance with the standard working curve.

3 Reagents and materials

Unless otherwise stated, the reagents used in this method are of analytical grade AND water is level II water as specified in GB/T 6682.

3.1 Reagents

3.1.1 Anhydrous ethanol (C₂H₆O).

3.1.2 Sodium hydroxide (NaOH).

3.1.3 Hydrochloric acid (HCl).

3.1.4 Citrate.

3.1.5 α-amylase: ≥ 1.5 U/mg.

3.1.6 Papain: ≥ 5 U/mg.

3.1.7 Sulfuric acid (H₂SO₄).

3.2 Reagent preparation

3.2.1 Ethanol solution (50%): MEASURE 500 mL of anhydrous ethanol; MIX it uniformly with 500 mL of water.

3.2.2 Sodium hydroxide solution (0.5 mol/L): WEIGH 20 g of sodium hydroxide; DISSOLVE it into 1000 mL of water; MIX it uniformly.

3.2.3 Sodium chloride solution (0.85%): WEIGH 8.5 g of sodium chloride; ADD water to dissolve and dilute it to 1000 mL; MIX it uniformly.

3.2.4 Hydrochloric acid solution (1 mol/L): ABSORB 83 mL of hydrochloric acid; USE water to dilute it to 1000 mL; MIX it uniformly.

3.2.5 Citrate buffer solution (pH 4.5): WEIGH 1.5 g of citric acid into a 100 mL beaker with a magnetic stirrer; ADD about 50 mL of distilled water to dissolve it; then ADD 12 mL of NaOH (1 mol/L); ADJUST the pH to 4.5 (with 0.1 mol/L HCl); TRANSFER the solution into a 100 mL volumetric flask; USE distilled water to make the volume reach to the mark. AND this buffer solution can be preserved for 3 d at 2 °C ~ 8 °C.

3.2.6 Protease-amylase solution: Respectively WEIGH 200 mg of papain and α -amylase; ADD 20 mL of water and GIND it to homogenate; CENTRIFUGE it at 3000 r/min for 5 min ~ 10 min; PREPARE it before use.

3.2.7 Sulfuric acid solution (3%): MEASURE 30 mL of sulfuric acid; ADD it to 1000 mL of water; MIX it uniformly.

3.3 Standard substance

Biotin (d-Biotin or Vitamin H) standard substance ($C_{10}H_{16}N_2O_3S$): purity $\geq 99\%$.

3.4 Standard solution preparation

3.4.1 Biotin standard stock solution (100 $\mu\text{g/mL}$): Accurately WEIGH 100 mg of biotin standard substance; USE ethanol solution (50%) to dissolve and transfer it into a 1000 mL volumetric flask; MAKE its volume reach to the mark. PRESERVE it in a brown bottle, which can be stored in a 2 °C ~ 4 °C refrigerator for 12 months.

3.4.2 Biotin standard intermediate solution (1.0 $\mu\text{g/mL}$): Accurately PIPETTE 1.00 mL of biotin standard stock solution into a 100 mL brown volumetric flask; USE ethanol solution (50%) to dissolve it and MAKE its volume reach to the mark. PRESERVE it in a brown bottle, which can be stored in a 2 °C ~ 4 °C refrigerator for 6 months.

3.4.3 Biotin standard working solution (10 ng/mL): Accurately PIPETTE 1.00 mL of biotin standard intermediate solution into a 100 mL volumetric flask; USE

machine; such samples as fruit and vegetable and semi-solid need to be mixed uniformly into homogenate; AND the liquid sample needs to be shaken before use to mix it, which is then preserved in a 4 °C refrigerator AND determined within 1 week.

6.2 Sample extraction

6.2.1 Potato, meat, dairy, fresh fruits and vegetables, algae samples, eggs, beans, nuts, animal offal and other natural foods: Accurately WEIGH an appropriate amount of homogeneous sample (m) (containing about 0.2 µg ~ 0.5 µg biotin), accurate to 0.001 g; PLACE it into a 50 mL volumetric flask; ADD 30 mL of citric acid buffer solution; SHAKE it and MAKE it subjected to high pressure hydrolysis at 121 °C for 15 min. TAKE out the sample; immediately COOL it to room temperature; ADD 1 mL of protease-amylase solution; PLACE it into a 36 °C ± 1 °C constant temperature incubator for incubation and enzymolysis for 16 h ~ 20 h; HEAT it in the 95 °C water batch for 30 min; then rapidly COOL it to room temperature; TRANSFER it into a 100 mL volumetric flask; USE water to make the volume reach to the mark (V₁).

6.2.2 Infant formula, cereals and other products (including raw and added biotin): Accurately WEIGH an appropriate amount of sample (m) (containing about 0.2 µg ~ 0.5 µg biotin), accurate to 0.001g; PLACE it into a 250 mL volumetric flask; ADD 100 mL of sulfuric acid solution; MAKE it subjected to hydrolysis at 121 °C for 30 min. After cooling, USE the sodium hydroxide solution to adjust pH to 4.5 ± 0.2; TRANSFER it into a 250 mL volumetric flask; USE water to make the volume reach to the mark; MIX it thoroughly. USE filter paper to filter it; DISCARD the first few milliliters; ABSORB 5 mL of the filtrate; ADD about 20 mL of water; USE sodium hydroxide solution to adjust the pH to 6.8 ± 0.2; TRANSFER it into a 100 mL volumetric flask; USE water to make the volume reach to the mark (V₁).

6.2.3 Strengthened biotin beverages or vitamin premixes and other samples: as for the liquid beverage, ADD 5 mL ~ 10 mL of sample to a 100 mL volumetric flask; ADD 50 mL of water; MIX it uniformly; TRANSFER it into a 100 mL volumetric flask; USE water to make the volume reach to the mark (V₁); as for the vitamin premixes: Accurately WEIGH appropriate amount of sample (m), accurate to 0.001 g; PLACE it into a 500 mL volumetric flask; ADD about 300 mL of water; MIX it uniformly. ADJUST the pH to 8.0 ± 0.2; TRANSFER it into a 1000 mL volumetric flask; USE water to make the volume reach to the mark (V₁).

6.3 Dilution

Appendix A

Medium and reagent

A.1 Lactobacillus agar medium

A.1.1 Component

TAKE 15.0 g of peptone milk, 5.0 g of yeast extract, 10.0 g of glucose, 100 mL of tomato juice, 2.0 g of potassium dihydrogen phosphate, and 1.0 g of polysorbate monooleate; ADD water to 1000 mL; ADJUST the pH to 6.8 ± 0.2 ($25\text{ }^{\circ}\text{C} \pm 5\text{ }^{\circ}\text{C}$).

A.1.2 Preparation method

ADD 10.0 g of agar into A.1.1; HEAT to boil it to dissolve the agar; after mixing it uniformly, CONTAIN it into different test tubes, 10 mL for each tube. STERILIZE it at $121\text{ }^{\circ}\text{C}$ under high pressure for 15 min; PREPARE for use.

A.2 Lactobacillus broth medium

A.2.1 Ingredients

TAKE 15.0 g of peptone milk, 5.0 g of yeast extract, 10.0 g of glucose, 100 mL of tomato juice, 2.0 g of potassium dihydrogen phosphate, and 1.0 g of polysorbate monooleate; ADD water to 1000 mL; ADJUST the pH to 6.8 ± 0.2 ($25\text{ }^{\circ}\text{C} \pm 5\text{ }^{\circ}\text{C}$).

A.2.2 Preparation method

HEAT the ingredients in A.2.1 to boil it; MIX it uniformly and CONTAIN it into different test tubes, 10 mL for each tube. STERILIZE it at $121\text{ }^{\circ}\text{C}$ under high pressure for 15 min

A.3 Biotin determination medium

A.3.1 Ingredients

TAKE 12.0 g of casein amino acid, 40.0 g of glucose, 20.0 g of sodium acetate, 0.2 g of L-cystine, 0.2 g of DL-tryptophane, 20.0 mg of adenine sulfate, 20.0 mg of guanine hydrochloride, 20.0 mg of uracil , 2.0 mg of thiamine hydrochloride, 2.0 mg of riboflavin, 2.0 mg of nicotinic acid, 2.0 mg of pantothenate, 4.0 mg of pyridoxine hydrochloride, 200.0 g of p-aminobenzoic acid, 1.0 g of dipotassium hydrogen phosphate, 1.0 g of magnesium sulfate,